Confounder Adjustment in Multiple Hypothesis Testing

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## Collaborators

### Confounder Adjustment

Qingyuan Zhao

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Model and Identifiability Estimation Hypothesis Tests

Numerical Examples

Summary

### Jingshu Wang



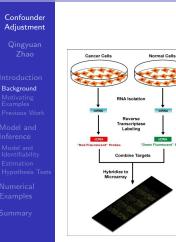
## Trevor Hastie



### Art Owen



## Microarray experiments



- Responses: normalized gene expression level.
- Primary variables (variables of interest): treatment, disease status, etc.
- Control covariates: age, gender, batch, date, etc.

## Microarray data analysis

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Biologist: "Which genes are (causally) related to this disease?" Statistician: "Let me run some analysis."

### Two common practices

- Sparse regression: regress the primary variable on the genes. More common for SNP data and predictive tasks.
- Association tests/screening (this talk): for each gene, perform a significance test of correlation with the primary variable.

Statistician: "Here a short list of candidate genes with false discovery rate (FDR)  $\leq 20\%$ ." Biologist: "Good, let me validate these discoveries."

## Concerns

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J. P. Ioannidis. Why most published research findings are false. *Chance*, 18(4):40–47, 2005

Two major challenges to reproducibility in genetic screening:

- **Correlated tests**: Is the FDR still controlled? If not, can we correct the statistical analysis?
  - Well studied in the last 15 years [Benjamini and Yekutieli, 2001, Storey et al., 2004, Efron, 2007, Fan et al., 2012].
- Confounded tests (this talk): the individual association tests are biased in presence of unobserved confounders. Can we still provide a good candidate list?
  - Equally long history [e.g. Alter et al., 2000, Price et al., 2006]. Still many open questions.

# Confounding

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### Brief history

- Fisher [1935] first uses the term in experiment designs.
- Kish [1959] first uses its modern meaning:
   A mixing of effects of unobserved extraneous factors (called confounders) with the effect of interest.
- Huge literature, but mostly in causal inference.

Aliases for confounders in genetic screening:

- "systematic ancestry differences" [Price et al., 2006].
- "batch effects" (widely used by biologists).
- "surrogate variables" [Leek and Storey, 2007, 2008].
- "unwanted variation" [Gagnon-Bartsch and Speed, 2012].
- "latent effects" [Sun et al., 2012].

# Example 1: gender study

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Which genes are more expressed in male/female?

A microarray experiment by Vawter et al. [2004]:

- Postmortem samples from the brains of 10 individuals.
- For each individual, 3 samples from different cortices.
- Each sample is sent to 3 different labs for analysis.
- Two different microarray platforms are used by the labs.

In total,  $10 \times 3 \times 3 = 90$  samples.

This example was first used by Gagnon-Bartsch and Speed [2012] to demonstrate the importance to "remove unwanted variation".

## Screening

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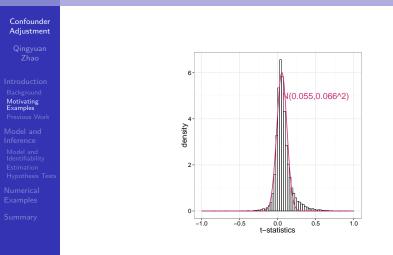
### Notation

- **Y**:  $n \times p$  matrix of gene expression.
- X:  $n \times 1$  vector of gender.
- Simplest association test:

Regress each column of Y (gene) on X.

- In R, run summary(lm(Y~X)).
- Equivalent to a two-sample *t*-test with equal variance.

# Histogram of t-statistics



Skewed and very underdispersed.

# What happened?

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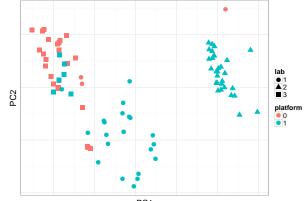
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PC1

## Association test

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### Notation

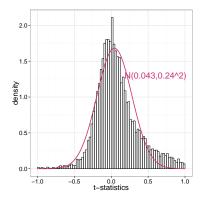
- **Y**:  $n \times p$  matrix of gene expression.
- X:  $n \times 1$  vector of gender.
- **Z**:  $n \times d$  matrix of control covariates (lab and platform).
- Modified association test:

Regress each column of Y (gene) on X and Z.

- In R, run summary(lm(Y~X+Z)).
- Report the significance of the coefficients of X.

## Histogram of t-statistics





Better, but still problematic. Reasonable guess: there are more unobserved confounders!

# Example 2: COPD study

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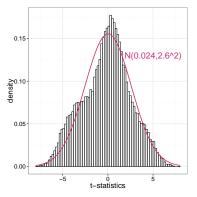
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- COPD = chronic obstructive pulmonary disease.
- Singh et al. [2011] tried to find genes associated with the severity of COPD (moderate or severe).



Overdispersed and skewed.

# Example 3: Mutual fund selection

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Barras et al. [2010] used the following model to select mutual funds:

$$Y_{it} = \alpha_i + \gamma_i^T \mathbf{Z}_t + e_{it}, \ i = 1, \dots, n, t = 1, \dots, p.$$

- $Y_{it}$ : observed log-return of fund *i* at time *t*.
- $\alpha_i$ : risk-adjusted return (Goal: find funds with positive  $\alpha$ ).
- **Z**<sub>t</sub>: systematic risk factors.

They assumed:

- $\alpha$  is sparse (Berk and Green equilibrium);
- No unobserved risk factors (is that possible/necessary?).

# Idea 0: Remove the largest principal component(s)

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### EIGENSTRAT [Price et al., 2006]

Regression model:

$$\mathbf{Y}_{n \times p} = \mathbf{X}_{n \times 1} \boldsymbol{\beta}_{p \times 1}^{T} + \mathbf{Z}_{n \times r} \boldsymbol{\Gamma}_{p \times r}^{T} + \mathbf{E}_{n \times p}$$

where **Z** is the first r PC(s) of **Y**.

- Motivation: in SNP, the largest PC(s) usually correspond to ancestry difference.
- Weakness: can easily remove true signals.

## Idea 1: Use control genes

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### Same regression model:

$$\mathbf{Y}_{n \times p} = \mathbf{X}_{n \times 1} \boldsymbol{\beta}_{p \times 1}^{T} + \mathbf{Z}_{n \times r} \boldsymbol{\Gamma}_{p \times r}^{T} + \mathbf{E}_{n \times p},$$

### RUV2 [Gagnon-Bartsch and Speed, 2012]

If we know  $oldsymbol{eta}_{\mathcal{C}}=oldsymbol{0}$  (negative controls),

- **1** Run PCA on  $col_{\mathcal{C}}(\mathbf{Y})$  to obtain **Z**.
- **2** Run the regression for  $col_{-C}(\mathbf{Y})$ .
  - Example: bacterial RNAs (spike-in controls).
  - Limited to the availability and number of negative controls.

# Idea 2: Sparsity

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Same regression model:

$$\mathbf{Y}_{n\times p} = \mathbf{X}_{n\times 1}\boldsymbol{\beta}_{p\times 1}^{T} + \mathbf{Z}_{n\times r}\boldsymbol{\Gamma}_{p\times r}^{T} + \mathbf{E}_{n\times p},$$

Idea: If  $\beta$  contains actual effects, it should be a sparse vector.

### SVA [Leek and Storey, 2008]

Iterate between

- Weighted PCA on **Y** (based on how likely  $\beta = 0$ ).
- Q Regress Y on X and the estimated PCs.
  - Does not always converge.

# Idea 2: Sparsity

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Same regression model:

$$\mathbf{Y}_{n\times p} = \mathbf{X}_{n\times 1}\boldsymbol{\beta}_{p\times 1}^{T} + \mathbf{Z}_{n\times r}\boldsymbol{\Gamma}_{p\times r}^{T} + \mathbf{E}_{n\times p},$$

Idea: If  $\beta$  contains actual effects, it should be a sparse vector.

### LEAPP [Sun, Zhang, and Owen, 2012]

**①** Run PCA on the residuals of  $\mathbf{Y} \sim \mathbf{X}$ .

2 Run a sparse regression.

# Our contributions: a unifying framework

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### Missing in previous methods:

- Explicit assumptions on the latent variables.
- Model identification conditions.
- Theoretical guarantees.
- Multiple primary and secondary covariates.
- Practical guidelines: when is confounder adjustment necessary/useful?

## Statistical model for confounding

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• Linear model for the responses (e.g. gene expression)

$$\mathbf{Y}_{n\times p} = \mathbf{X}_{n\times 1} \boldsymbol{\beta}_{p\times 1}^{T} + \mathbf{Z}_{n\times r} \boldsymbol{\Gamma}_{p\times r}^{T} + \mathbf{E}_{n\times p},$$

- X: primary variable (disease, treatment, gender, etc.);
- Z: unobserved confounders;
- $\beta$ : primary effects that we are interested in.
- Missing in the literature: dependence of Z and X

$$\mathbf{Z}_{n\times r} = \mathbf{X}_{n\times 1}\boldsymbol{\alpha}_{r\times 1}^{T} + \mathbf{W}_{n\times r},$$

• Additional distributional assumptions:  $X_i \stackrel{\text{i.i.d.}}{\sim} \text{mean } 0, \text{ variance } 1, i = 1, ..., n,$   $\mathbf{E} \stackrel{\text{i.i.d.}}{\sim} N(\mathbf{0}, \Sigma), \mathbf{E} \perp (\mathbf{X}, \mathbf{Z}), \Sigma = \text{diag}(\{\sigma_j^2\}_{j=1}^p),$  $\mathbf{W} \stackrel{\text{i.i.d.}}{\sim} N(\mathbf{0}, \mathbf{I}_r), \mathbf{W} \perp \mathbf{X}.$ 

## Marginal effects and direct effects

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### The model can be rewritten as

$$\mathbf{Y}_{n\times p} = \mathbf{X}_{n\times 1} \left( \beta_{p\times 1} + \Gamma_{p\times r} \alpha_{r\times 1} \right)^T + (\mathbf{W}\Gamma + \mathbf{E}),$$

which gives the population identity

$$au_{p imes 1} = eta + \Gamma lpha.$$

- τ: marginal effects.
- $\beta$ : direct effects (more meaningful).

# COPD data: marginal effects vs. direct effects

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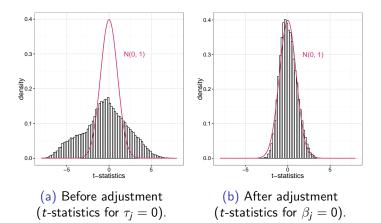
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# Identifiability of ${\boldsymbol{\beta}}$

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## To identify lpha and eta from

$$\tau_{p\times 1} = \beta_{p\times 1} + \Gamma \alpha_{r\times 1},$$

there are p equations but p + r parameters.

### Proposition [Wang, Z., Hastie, and Owen, 2015]

Suppose  $\Gamma$  can be identified.  $\beta$  is identifiable under either of the two following conditions:

Negative control: for a known negative control set C,

 $\boldsymbol{\beta}_{\mathcal{C}} = \mathbf{0}, \ |\mathcal{C}| \geq r, \ \mathrm{rank}(\boldsymbol{\Gamma}_{\mathcal{C}}) = r.$ 

**2** Sparsity:  $\|\beta\|_0 \leq \lfloor (p-r)/2 \rfloor$  (the maximum breakdown point),

 $\operatorname{rank}(\Gamma_{\mathcal{C}}) = r, \ \forall \mathcal{C} \subset \{1, \dots, p\} \ \mathrm{such \ that} \ |\mathcal{C}| = r.$ 

## Rotation

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### Householder transformation

$$\mathbf{X}_{n\times 1} = \mathbf{Q}\mathbf{R}$$

where  $\mathbf{Q} \in \mathbb{R}^{n \times n}$  is orthogonal with  $\mathbf{R} = (\|\mathbf{X}\|_2, 0, \dots, 0)^T$ . • For simplicity, assume  $\|\mathbf{X}\|_2 = \sqrt{n}$ .

• Can be easily extended to multiple variables **X**.

### Rotation (LEAPP)

Left-Multiply 
$$\mathbf{Q}^T$$
 to  $\mathbf{Y} = \mathbf{X}\boldsymbol{\beta}^T + \mathbf{Z}\Gamma^T + \mathbf{E}$ , we get  
 $\operatorname{row}_1(\mathbf{Q}^T\mathbf{Y}) \sim \operatorname{N}(\sqrt{n}(\boldsymbol{\beta} + \Gamma\boldsymbol{\alpha}), \Gamma\Gamma^T + \boldsymbol{\Sigma}),$   
 $\operatorname{row}_{-1}(\mathbf{Q}^T\mathbf{Y}) \stackrel{\text{i.i.d.}}{\sim} \operatorname{N}(\mathbf{0}, \Gamma\Gamma^T + \boldsymbol{\Sigma}).$ 

## Two-step estimation

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## Run factor analysis for

$$\operatorname{row}_{\text{-}1}(\boldsymbol{\mathsf{Q}}^{\mathcal{T}}\boldsymbol{\mathsf{Y}}) \overset{\text{i.i.d.}}{\sim} \operatorname{N}(\boldsymbol{\mathsf{0}},\boldsymbol{\Gamma}\boldsymbol{\Gamma}^{\mathcal{T}}+\boldsymbol{\Sigma})$$

to obtain Γ̂ and Σ̂. Identifiability follows from classical results in factor analysis [e.g. Anderson and Rubin, 1956].
Run linear regression for the marginal effects

$$\frac{\operatorname{row}_{1}(\mathbf{Q}^{T}\mathbf{Y})_{p\times 1}}{\sqrt{n}} = \hat{\mathbf{\Gamma}}_{p\times r} \alpha_{r\times 1} + \beta_{p\times 1} + \tilde{\mathbf{E}}_{1}/\sqrt{n}$$
response design matrix coefficients

# How accurate is $\hat{\Gamma}?$

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### Assumptions

- High-dimensional data:  $n \to \infty$ ,  $p \to \infty$ .
- Assume that the factors are strong enough:  $\lim_{p \to \infty} \frac{1}{p} \Gamma^T \Sigma^{-1} \Gamma \text{ exists and is positive definite.}$
- Consistent estimate of r [Bai and Ng, 2002].

### Theoretical Results for MLE

 $\bullet\,$  Consistent estimate of  $\Gamma$  and  $\Sigma$  [Bai and Li, 2012] and

$$\sqrt{n}(\hat{\boldsymbol{\Gamma}}_j - \boldsymbol{\Gamma}_j) \stackrel{d}{\rightarrow} \mathrm{N}(\boldsymbol{0}, \sigma_j^2 \boldsymbol{\mathsf{I}}_r), \ \sqrt{n}(\hat{\sigma}_j - \sigma_j) \stackrel{d}{\rightarrow} \mathrm{N}(\boldsymbol{0}, 2\sigma_j^4),$$

• Uniform consistency if  $n^k/p \to \infty$  for some k > 0 [Wang, Z., Hastie, and Owen, 2015].

# Strategy 1: Estimate $\beta$ via negative controls

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## Recall the marginal effects are

$$\tilde{\mathbf{Y}}_{p\times 1}^{T} = \mathbf{\Gamma}_{p\times r} \, \boldsymbol{\alpha}_{r\times 1} + \boldsymbol{\beta}_{p\times 1} + \tilde{\mathbf{E}}_{1}/\sqrt{n}$$
**response design matrix coefficients**
In the negative control scenario, we know  $\boldsymbol{\beta}_{\mathcal{C}} = \mathbf{0}$ .

### Generalized Least Squares (GLS) estimator

$$\hat{oldsymbol{lpha}}^{\mathrm{NC}} = (\hat{oldsymbol{\Gamma}}_{\mathcal{C}}^{T} \hat{\Sigma}_{\mathcal{C}}^{-1} \hat{oldsymbol{\Gamma}}_{\mathcal{C}})^{-1} \hat{oldsymbol{\Gamma}}_{\mathcal{C}}^{T} \hat{\Sigma}_{\mathcal{C}}^{-1} \tilde{oldsymbol{Y}}_{1,\mathcal{C}}^{T} / \|oldsymbol{X}\|_{2}$$
 $\hat{eta}_{\mathcal{C}}^{\mathrm{NC}} = ilde{oldsymbol{Y}}_{1,\mathcal{C}}^{T} / \|oldsymbol{X}\|_{2} - \hat{oldsymbol{\Gamma}}_{\mathcal{C}} \hat{oldsymbol{lpha}}^{\mathrm{NC}}$ 

Note: RUV4 [Gagnon-Bartsch et al., 2013] = Ordinary Least Squares (OLS).

# Asymptotic distribution of $\hat{oldsymbol{eta}}^{ ext{NC}}$

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### Theorem (Wang, Z., Hastie, and Owen [2015])

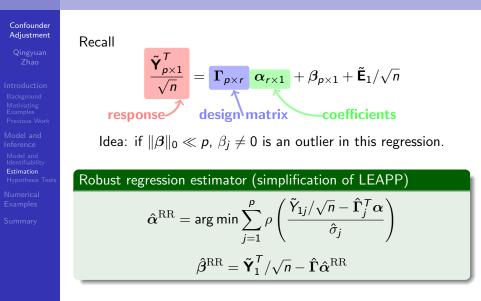
Under the assumptions of uniform convergence of  $\hat{\Sigma}$  and  $\hat{\Gamma}$  and  $\lim_{p \to \infty} \frac{1}{|\mathcal{C}|} \Gamma_{\mathcal{C}}^T \Sigma_{\mathcal{C}}^{-1} \Gamma_{\mathcal{C}} \succ \mathbf{0}$ , then for any finite index set S such that  $S \cap \mathcal{C} = \emptyset$ :

• If the number of negative controls  $|\mathcal{C}| \to \infty$ ,

$$\sqrt{n}(\hat{eta}^{ ext{NC}}_{\mathcal{S}}-eta_{\mathcal{S}}) \stackrel{d}{
ightarrow} ext{N}(m{0},(1+\|m{lpha}\|_2^2)m{\Sigma}_{\mathcal{S}})$$

$$\begin{array}{l} \textit{If } \lim_{p \to \infty} |\mathcal{C}| < \infty, \\ & \sqrt{n} (\hat{\beta}^{\mathrm{NC}}_{\mathcal{S}} - \beta_{\mathcal{S}}) \stackrel{d}{\to} \mathrm{N}(\mathbf{0}, (1 + \|\boldsymbol{\alpha}\|_2^2) (\boldsymbol{\Sigma}_{\mathcal{S}} + \boldsymbol{\Delta}_{\mathcal{S}})) \\ & \textit{where } \boldsymbol{\Delta}_{\mathcal{S}} = \lim_{p \to \infty} \Gamma_{\mathcal{S}} (\Gamma_{\mathcal{C}}^T \boldsymbol{\Sigma}_{\mathcal{C}}^{-1} \Gamma_{\mathcal{C}})^{-1} \Gamma_{\mathcal{S}}^T. \end{array}$$

# Strategy 2: Estimate $\beta$ via sparsity



# Asymptotic distribution of $\hat{oldsymbol{eta}}^{\mathrm{RR}}$

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### Assumptions on the loss function $\rho(x)$

The derivatives  $\rho'$ ,  $\rho''$  and  $\rho'''$  exist and are bounded.  $\rho(0) = \rho'(0) = 0$ ,  $\rho''(0) > 0$  and  $\rho'(x) \cdot x \ge 0$ . (e.g. Tukey's bisquare)

### Theorem (Wang, Z., Hastie, and Owen [2015])

Under the assumptions of uniform convergence of  $\hat{\Sigma}$  and  $\hat{\Gamma}$  and the above assumption of the loss function, if  $\min(\|\beta\|_0, \|\beta\|_1)\sqrt{n/p} \to 0$ , then for any finite index set S:

 $\sqrt{n}(\hat{eta}_{\mathcal{S}}^{\mathrm{RR}} - eta_{\mathcal{S}}) \stackrel{d}{
ightarrow} \mathrm{N}(\mathbf{0}, (1 + \|oldsymbol{lpha}\|_2^2) \mathbf{\Sigma}_{\mathcal{S}}).$ 

# Oracle efficiency

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In either the sparsity or negative control scenario (  $|\mathcal{C}| \to \infty)$  :

$$\sqrt{n}(\hat{eta}_{\mathcal{S}} - eta_{\mathcal{S}}) \stackrel{d}{
ightarrow} \mathrm{N}(\mathbf{0}, (1 + \|oldsymbol{lpha}\|_2^2) \mathbf{\Sigma}_{\mathcal{S}})$$

### Oracle estimator

Consider the model

$$\mathbf{Y} = \mathbf{X}\boldsymbol{\beta}^{\mathsf{T}} + \mathbf{Z}\boldsymbol{\Gamma}^{\mathsf{T}} + \mathbf{E}.$$

If **Z** were observed, the oracle OLS estimator would be  $\sqrt{n}(\hat{\beta}_{S}^{\mathrm{OLS}} - \beta_{S}) \sim \mathrm{N}(\mathbf{0}, (1 + \|\boldsymbol{\alpha}\|_{2}^{2})\boldsymbol{\Sigma}_{S}).$ 

 $\hat{oldsymbol{eta}}_{\mathcal{S}}$  is as efficient asymptotically as the oracle estimator!

# Significance test for confounding

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### Theorem (Wang, Z., Hastie, and Owen [2015])

Under the above assumptions for oracle efficiency and the null hypothesis that  $H_{0,\alpha}$ :  $\alpha = \mathbf{0}$ , we have

$$\mathbf{n}\cdot\hat{\boldsymbol{\alpha}}^{\mathsf{T}}\hat{\boldsymbol{\alpha}}\overset{d}{\rightarrow}\chi_{\mathsf{r}}^{2}$$

where  $\chi_r^2$  is the chi-square distribution with r degree of freedom.

### Recipes

- Graphical diagnostics: the histogram of test statistics.
- Positive controls: e.g. X/Y genes for gender.
- $\ensuremath{\mathfrak{S}} \ensuremath{\mathfrak{S}} \ensurema$

## Multiple hypothesis testing

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### Two-sided asymptotic z-tests

Test 
$$H_{j0}$$
:  $\beta_j = 0$  vs.  $H_{j1}$ :  $\beta_j \neq 0$  for  $j = 1, \dots, p$ .

$$t_j = rac{\sqrt{neta_j}}{\hat{\sigma}_j \sqrt{1+\|\hat{lpha}\|^2}}, \; P_j = 2(1-\Phi(|t_j|)).$$

### Theorem (Wang, Z., Hastie, and Owen [2015])

Under the assumptions for oracle efficiency, the overall type I error and the familywise error rate (FWER) can be asymptotically controlled.

FDR control: ongoing work.

## Simulation: n = 100, p = 5000 and r = 10

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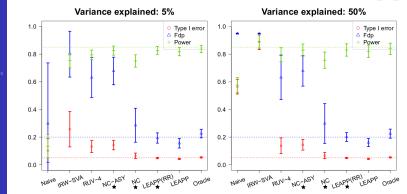
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- Sparsity:  $\|\beta\|_0/p = 0.05$ ; NC:  $|\mathcal{C}| = 30$ .
- $\Gamma$  uniform from orthogonal matrices;  $\sigma_i^2 \stackrel{i.i.d.}{\sim} \text{InvGamma}(3,2).$
- Variance of **X** explained by **Z**:  $\max_{\rho} \operatorname{corr}(X_i, \rho^T \mathbf{Z}_i) = \frac{\|\boldsymbol{\alpha}\|_2}{1 + \|\boldsymbol{\alpha}\|_2}$



## COPD data: severity as primary variable

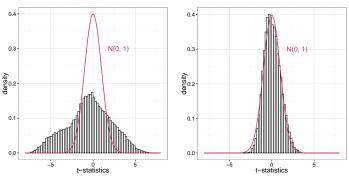


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(a) Naive linear regression.

(b) After adjustment.

- $\hat{r} = 1$  [Onatski, 2010].
- $\hat{\alpha} \approx$  0.98, variance explained is approximately 22%.
- Test of confounding: p-value  $\approx$  0.

# COPD data: gender as primary variable

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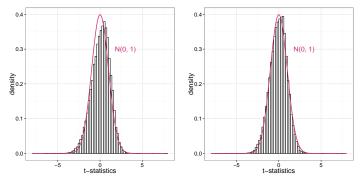
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Genes associated with gender should come from X/Y chromosomes (positive controls).

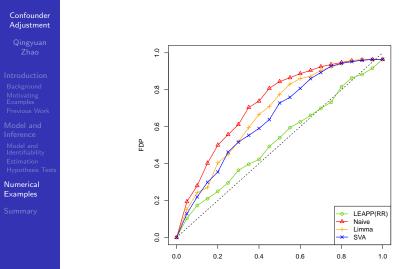


(a) Naive linear regression.

(b) After adjustment.

- $\hat{\alpha} \approx -0.27$ , variance explained is approximately 3%.
- Test of confounding: p-value  $\approx 1.2 \times 10^{-3}$ .

## COPD data: gender as primary variable



Nominal FDR

# COPD data: gender as primary variable



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Method	X/Y Genes in Top 100
LEAPP(RR)	69
Naive	58
Limma	58
SVA	68

# Mutual fund selection (preliminary results)

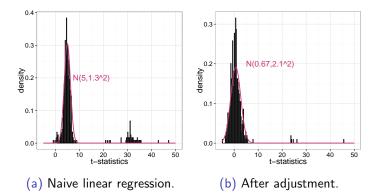
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- p = 469 mutual funds with monthly returns available in CRSP database in Jan. 1980 - Dec. 2000 (n = 240).
- Apply the RR procedure with r = 6 without adjusting for any observed systematic risk factor.



# Summary

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Model and Identifiability Estimation Hypothesis Test

Numerical Examples

Summary

### Recap

- Linear model with unobserved confounding factors.
- Identification conditions: negative control and sparsity.
- Two-step estimation of the primary effects.
- Asymptotic distributions and oracle efficiency.
- Hypothesis tests for confounding and the primary effects.

### Open problems

- Correlated noise: approximate factor models.
- Weak factors: random matrix theory.
- Non-Gaussian data: RNA-seq, GWAS.
- Beyond linearity?

## Resources

#### Confounder Adjustment

Qingyuan Zhao

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Summary

- J. Wang, Z., T. Hastie, and A. B. Owen. Confounder adjustment in multiple hypothesis testing. *under revision for Annals of Statistics*, 2015.
  - Available on arXiv.
- Software: cate on CRAN.

(https://cran.r-project.org/web/packages/cate/index.html)

- Package vignette available online.
- Unified interface for existing packages sva, ruv, leapp.
- We also support formula:

results <- cate( $\sim$  gender | . - gender - 1, data, ...)